

THE MELOSATINS—A NOVEL CLASS OF ALKALOIDS FROM *MELOCHIA TOMENTOSA*^a

G. J. KAPADIA*, Y. N. SHUKLA and S. P. BASAK

Department of Biomedical Chemistry, College of Pharmacy and Pharmacal Sciences, Howard University,
Washington, DC 20059, U.S.A.

E. A. SOKOLOSKI and H. M. FALES*

Laboratory of Chemistry, National Heart, Lung, and Blood Institute National Institutes of Health, Bethesda,
MD 20205, U.S.A.

(Received U.S.A. 3 December 1979)

Abstract—Details of the isolation of melosatin A, B, and C and the synthesis of melosatin A are presented. Melosatin C has been characterized as 7-methoxy-4-(5-phenylpentyl)isatin. Several Me and OMe substituted isatins are synthesized as models and UV and mass spectra of the series are discussed.

RECENTLY we reported the isolation and structures of several of the constituents of the tumorigenic plant, *Melochia tomentosa* L. (Sterculiaceae), including an unusual quinolinone alkaloid, melochinone (1)¹, its open chain analog, melovinone (2)², 6-methoxy-7,8-methylenedioxy coumarin (3)³, the cyclopeptide alkaloids, melonovines A (4) and B (5), and scutianine B (6).⁴ We have also communicated on two novel isatin alkaloids melosatin A and B⁵ (7 and 8 respectively) from the same source. This paper details further the structure of these compounds and reports the isolation and characterization of another alkaloid in this series, melosatin C.

As reported earlier, melosatin A and B (7 and 8 respectively) are yellow alkaloids giving the bluish green color reaction with sulfuric acid typical of isatins.⁶ Melosatin A⁷ also reduces to an alcohol with sodium borohydride, undergoes ring expansion⁶ to quinolines 10 and 11, forms a methoxime, and condenses with *o*-phenylenediamine to adducts 12 and 13; details are reported below. While the structure of the unsubstituted melosatin B (8) was unequivocally determined from its ¹H NMR spectrum,⁵ the NMR of melosatin A was in accord both with 7 and an isomer having the two methoxyls located in the 5 and 6 positions of the isatin ring. Because 5-nitrovanillin was more readily available than intermediates leading to 2,3-dimethoxy-5-nitrobenzaldehyde, isomer 7 was synthesized. Thus 5-nitrovanillin was methylated with diazomethane producing methyl ether (14) rather than the reported acetophenone.⁸ This was condensed with benzalacetone (15) and the resulting substituted dibenzalacetone (16) reduced to an amorphous mixture of saturated and unsaturated alcohols with sodium borohydride. Without further purification, the remaining olefinic bonds as well as the nitro group were reduced with Pd-C yielding a crystalline, saturated amino alcohol (17).⁹ This was dehydrated with P₂O₅, forming a mixture of olefins which were reduced directly to the desired aniline (18) which was cyclized with oxalyl chloride affording a product, isolated by tlc,

identical in all respects (m.p., m.m.p., IR, UV, NMR, MS) with melosatin A, establishing its structure as 7.

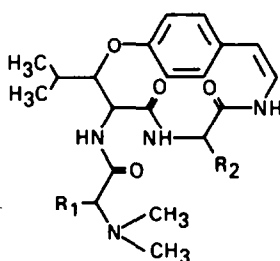
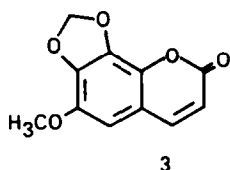
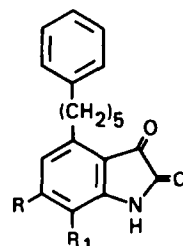
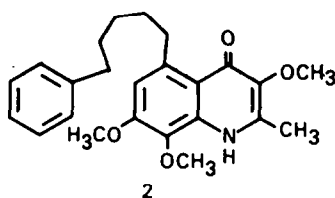
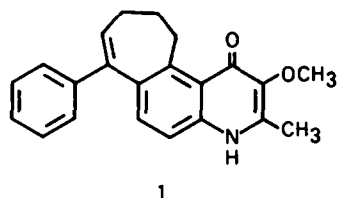
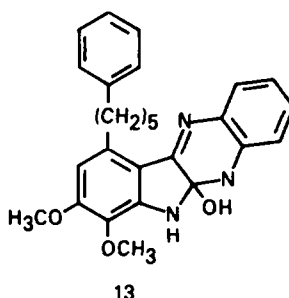
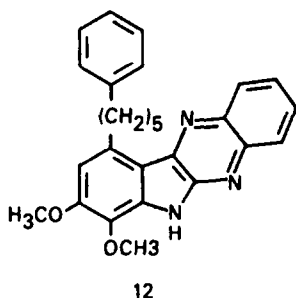
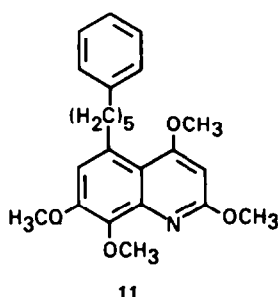
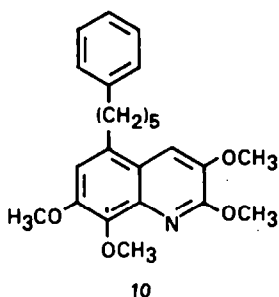
Melosatin C is a red alkaloid isolated from the same crude product as melosatin A and B. Its IR, mass, and ¹H NMR spectra are very similar to those of melosatin A and B, but its formula (C₂₀H₂₁NO₃) and ¹H NMR permit a single OMe group. This group is located on the isatin nucleus as proved both by the presence of five phenyl protons in the ¹H NMR (δ 7.26) and by the masses of the benzyl ion (*m/z* 91) and isatin fragment (*m/z* 232) in its mass spectrum (see below).

The five carbon side chain was easily located at C-4 as proved by a pronounced downfield shift of the benzylic methylene group at C₄ (δ 2.90) caused by the *peri* carbonyl at C₃. This effect has been observed earlier with melosatin A and B⁵ and melochinone (1).¹ In the phenylenediamine adduct of melosatin A (12) the effect was especially pronounced (δ 3.50), undoubtedly due to the influence of the newly formed phenazine ring.

Location of the aromatic OMe on the isatin ring was less straightforward. Two vicinal aromatic protons are present at δ 6.84 and δ 7.06. Using melosatin A, B and N-methylisatin as models,⁵ the latter is approximately the value expected for a C-6 proton shifted upfield 0.36 ppm by an adjacent OMe group. The remaining proton can be located either at C-5 or C-7. The former was favored (structure 9) because of its relatively broad signal which we ascribe to coupling with the adjacent benzylic methylene group of the side chain. To confirm this feature, 4-methyl-7-methoxyisatin was synthesized from the readily available 2-methoxy-5-methylaniline via the Sandmeyer reaction.¹⁰ The chemical shifts of the aromatic protons of the product agree closely with those of melosatin C [δ 6.79(d, J = 8.5 Hz), δ 7.04(d, J = 8.5 Hz) and δ 6.84(d, J = 8 Hz), δ 7.06(d, J = 8 Hz), respectively]. Furthermore, the upfield doublet (H-5) of the model was indeed broadened by coupling to the adjacent 4-Me group. The UV spectra of both also agree (Fig. 2) and melosatin C is assigned structure 9.

The colors of the melosatin and isatin range from light yellow to dark red as indicated by variation of the long wavelength absorption tail (Figs 1 and 2) arising from the *n* → π^* transition of the dicarbonyl system. The structural features responsible for this variation are not

^aPart 13 in the series, Potential Carcinogens. For Part 12 see Ref. 2.

7. R = R₁ = OCH₃8. R = R₁ = H9. R = H; R₁ = OCH₃4. R₁ = CH(CH₃)₂, R₂ = CH₂CH(CH₃)₂5. R₁ = CH(CH₃)₂, R₂ = CH₂C₆H₄OH6. R₁ = R₂ = CH₂C₆H₅

entirely clear from this series, but it appears that a single substituent (alkyl or OMe) in the 4-position of the isatin nucleus causes a shift to yellow. Attachment of a second OMe in the 7-position offsets this hypsochromic effect unless the OMe is forced out of the plane of the ring by yet a third substituent in the 6-position.

The mass spectra of melosatins A-C exhibit ions for loss of 1 and 2 moles of CO as expected for isatins, ions at m/z 91 and $M^+ - 91$ from cleavage of the benzyl groups. Important ions, supported by metastables, are observed at $[M-131]^+$ which apparently arise via transfer of hydrogens from the pentyl side chain to locations on the isatin ring via a process such as a-b:

$[M-131]^+$ ions then lose the isatin carbonyls in rapid sequence (one metastable ion for C₂H₂O₂) in an unusual

reversion to an odd-electron ion represented above as c or d.

The ions at $(M-131)^+$ are always accompanied by ions at $(M-132)^+$ involving transfer of only one hydrogen that may arise from the process e-f.

The mass spectra of the isatins synthesized in this study are unexceptional (Experimental); all show good molecular ions with loss of CO taking precedence over all other fragmentations. This is followed by loss of HCN in the case of isatin, 4-methylisatin, and 4,7-dimethoxyisatin. In all other cases, loss of the combination of methyl and CO, in either sequence, is an important process.

Indole and oxindole alkaloids are widely distributed in nature but to our knowledge this is the first recorded

RED ISATINS

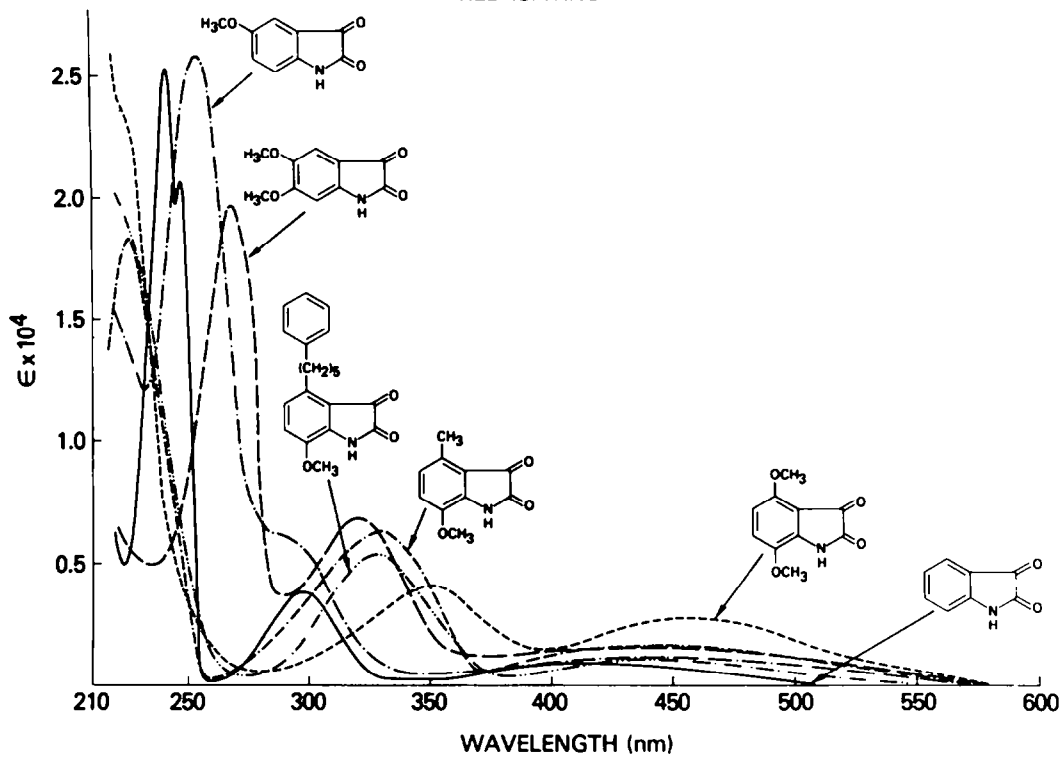


Fig. 1.

YELLOW ISATINS

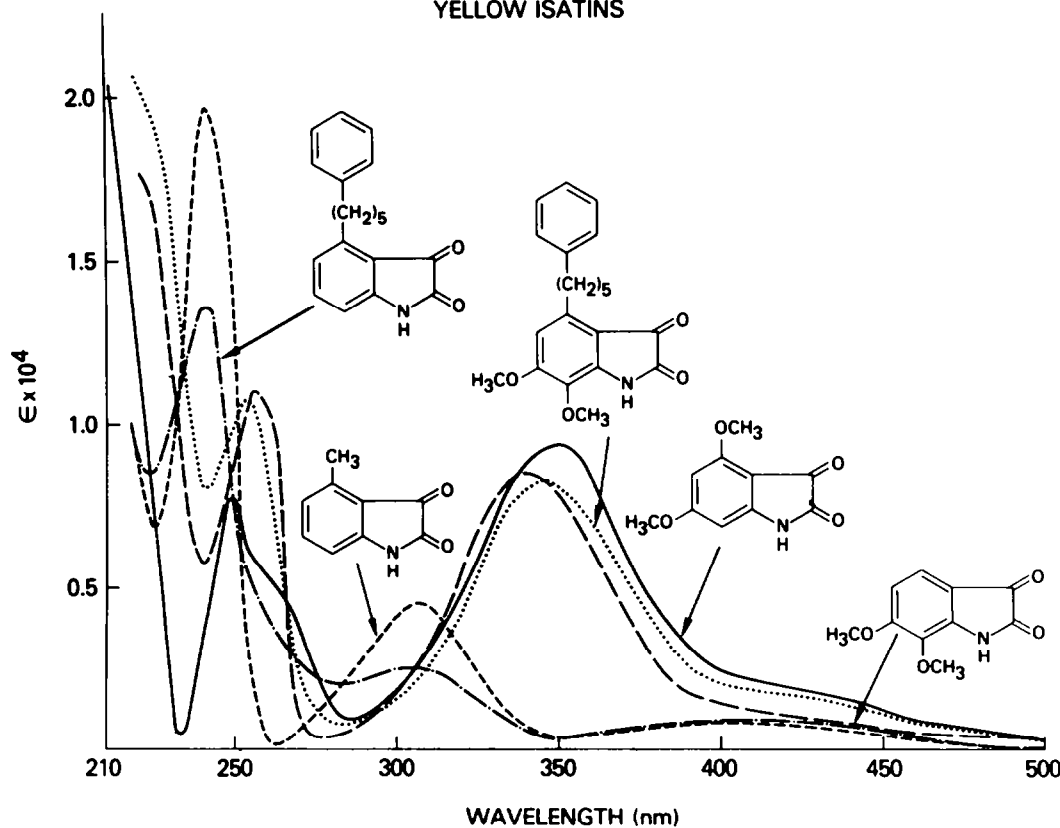
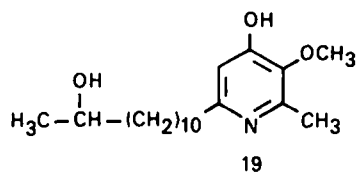
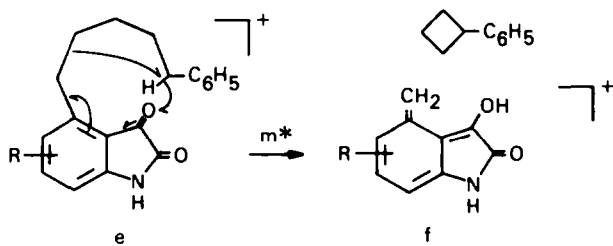
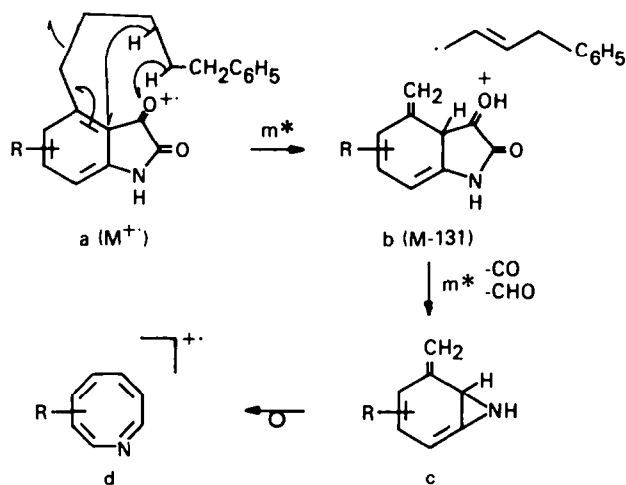
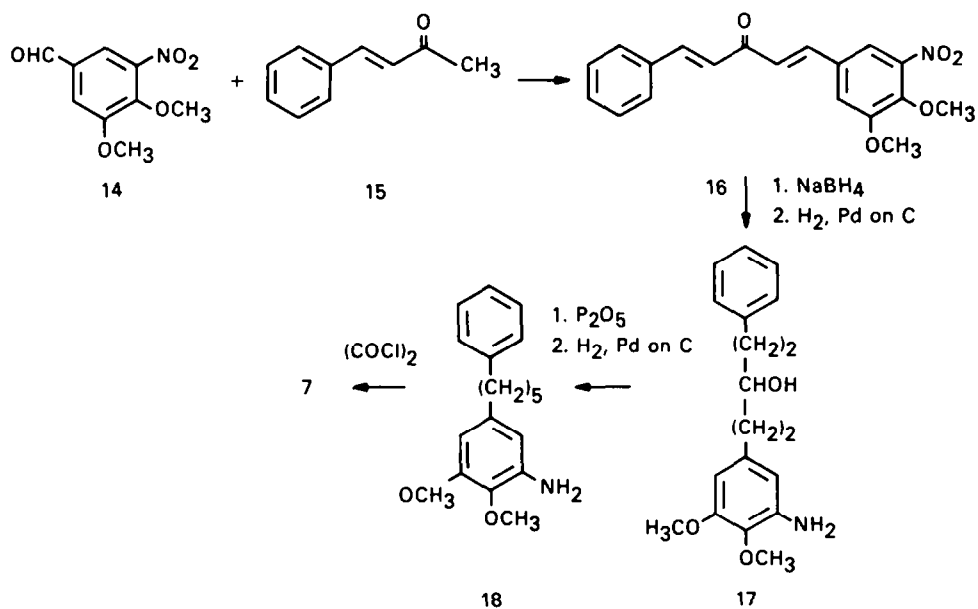


Fig. 2.



instance where they have reached the oxidation state of isatin. The isatins are unlikely to be artifacts, however, since their presence is easily observed by tlc prior to extensive work-up.

Melosatin A appears to be closely related to melovinine (2), except that the hypothetical anthranilate-pyruvate construction of the quinolinone ring¹¹ has been replaced by anthranilate-formate. Alternatively, *o*-aminophenylpyruvate from degradation of kynurenine could be the source of this ring. Melosatin B is similarly related to melochinone (1) except that cyclization of the 5-carbon chain to a 7-membered ring has not occurred.

Recently, Medina and Spittler¹² have reported the isolation of a pyridone alkaloid, melochinin (19) from the closely related *Melochia pyramidata* L. (Sterculiaceae). Although it bears some resemblance to the quinolinone alkaloids melochinone (1) and melovinine (2) insofar as the substitution of the pyridone ring is concerned, its biogenesis appears to be quite different.

EXPERIMENTAL

M. ps were determined on a Thomas-Hoover capillary melting point apparatus and are uncorrected. IR spectra were determined in nujol and UV spectra in EtOH. 100 MHz ¹H NMR spectra were determined on a Varian XL-100 spectrometer equipped with a Digilab Fourier Transform data system in CDCl₃ with TMS as internal reference. All tlc was carried out on Silica gel-60 precoated plates, F-254 (E. Merck). Visualization of tlc was effected by short and long-wave UV and 2,4-dinitrophenylhydrazine and Dragendorff sprays. 5-Methoxyisatin, isatin, and 5,6-dimethoxyisatin were purchased from K&K Chemical Co.

A voucher specimen, identified by Dr. Julia Morton, has been deposited in the Morton Collectanea, Univ. of Miami, Coral Gables, Florida.

Extraction and isolation. Dried and ground roots (1.6 kg) of *M. tomentosa* (collected at Curacao by Mr. W. P. Maal) were extracted in turn with boiling petroleum ether, benzene, and methylene chloride (2 days, 5 l each solvent). The residual material was combined with Ba(OH)₂ (50 g) and 50% aqueous EtOH (1.2 l), air dried and then continuously extracted with CH₂Cl₂ (2 days, 5 l). The benzene extract and the first CH₂Cl₂ extract were identical on tlc and combined (extract A) while the later CH₂Cl₂ extract was labelled B.

Melosatin A (7). The residue (7 g) obtained by the removal of solvent from extract A was chromatographed on silica gel (100 g). The column was eluted with increasingly polar mixtures of petroleum ether, benzene, and chloroform. The fractions obtained in benzene-chloroform (3:1) were combined and further purified by preparative tlc in benzene EtOH (3:1). The yellow residue isolated from the adsorbent at *R_f* 0.5 was crystallized from benzene-petroleum ether to give 16 mg (0.001%) of yellow melosatin A (7): m.p. 119–21°; IR (nujol) ν_{\max} 3200 (NH), 1750, 1725 (CO), 1645, (CONH), 1250 and 1135 cm⁻¹ (OMe); MS *m/z* 353.162 (100, M⁺. C₂₁H₂₃NO₄ requires 353.1626), 262 (13), 222 (35), 221 (40), 91 (98); for ¹H NMR see Ref. 5.

Melosatin B (8). The residue (4 g) obtained by the removal of solvent from extract B was chromatographed over silica gel (300 g). The column was eluted with solvents of increasing polarity. A viscous material was obtained with benzene-chloroform (1:3) which was purified by preparative tlc using CHCl₃-MeOH (95:5) to give 4 mg (0.00025%) of yellow amorphous melosatin B: MS *m/z* 293 (95, M⁺. C₁₉H₁₉NO₂), 202 (57), 162 (80), 161 (84), 133 (47), 91 (100); for ¹H NMR see Ref. 5.

Melosatin C (9). During the chromatography of extract B, a red material was obtained from the chloroform-benzene (1:1) eluate. It was purified by preparative tlc using CHCl₃-MeOH

(95:5) (*R_f* 0.67) and crystallized from benzene-petroleum ether providing 5 mg (0.0003%) of red needles of melosatin C: m.p. 124–5°; IR (nujol) ν_{\max} 3200 (NH), 1745, 1730 (CO), 1635 (CONH), 1595, 1515, 1242 and 1155 cm⁻¹; MS *m/z* 323 (100, M⁺. C₂₀H₂₁NO₃), 232 (33), 192 (60), 191 (47), 176 (25), 162 (21), 135 (72), 91 (88); ¹H NMR δ 1.56 (broad m, CCH₂, 6 H), 2.62 (t, C₆H₄CH₂, 2 H), 2.90 (t, isatin CH₂, 2 H), 3.90 (s, OMe, 3 H), 6.84 (d, *J* = 8 Hz, 5 H of isatin, 1 H), 7.06 (d, *J* = 8 Hz, 6 H of isatin, 1 H), 7.24 (m, C₆H₅, 5 H), 7.68 (broad s, NH, 1 H).

***o*-Phenylenediamine adduct (12) of melosatin A.** Melosatin A (2 mg) and *o*-phenylenediamine (4 mg) in AcOH (1 ml) were refluxed on the water bath for 2 hr. At the end of the reaction, the solvent was removed under N₂. The residue obtained was subjected to preparative tlc in CHCl₃-EtOAc (3:1) and two yellow products were obtained. The most mobile, *R_f* 0.35, is the adduct 12: MS 425 (40, M⁺. C₂₇H₂₇N₃O₂), 334 (25), 320 (25), 306 (53), 293 (100), 278 (46), 91 (34); ¹H NMR δ 1.55 (broad m, C(CH₂)₃C, 6 H), 2.70 (t, C₆H₅CH₂, 2 H), 3.50 (t, isatin CH₂, 2 H), 4.05 (s, OMe, 6 H), 6.77 (s, isatin 5 H, 1 H), 7.25 (m, C₆H₅, 5 H), NH too broad to observe. The less mobile spot is assumed to be the incompletely dehydrated adduct 13 on the basis of its mass spectrum: *m/z* 443 (20, M⁺. C₂₇H₂₅N₃O₃), 425 (45, M⁺ - H₂O), 334 (24), 320 (25), 310 (100), 306 (45), 293 (80), 278 (40), 91 (32).

Treatment of melosatin A (7) with diazomethane. Melosatin A (2 mg) was dissolved in an excess of ethereal diazomethane (10 ml) and the mixture was allowed to stand overnight at room temp. The solvent was removed and the residue subjected to GC-MS. Two isomeric methoxyquinolines eluted, one at 270° and one at 291° when programmed at 10°/min on a 1% OV-17 column. The two are assumed to be structures 10 and 11 on the basis of their molecular ions and the known reactions of isatins with diazomethane.⁶ The early-eluting isomer shows far more loss of Me than the second, but differentiation between their structures on this basis is not obvious: MS (isomer eluting at 270°) 395 (100), M⁺. C₂₄H₂₉NO₄, 380 (59), 366 (34), 262 (49), 91 (10); MS (isomer eluting at 291°) 395 (100, M⁺. C₂₄H₂₉NO₄), 380 (13), 366 (7), 262 (15), 91 (7).

Melosatin A methoxime. A submilligram quantity of melosatin A was allowed to stand in pyridine overnight with an excess of methoxyamine hydrochloride. The solvents were evaporated under a stream of N₂ and the residue extracted into CHCl₃ from added water. Upon evaporation, the residue was applied directly to the direct insertion probe of the mass spectrometer: MS N₂ 382 (45, M⁺. C₂₂H₂₆ m/z O₄), 351 (100), 335 (35), 91 (45).

Sodium borohydride reduction of melosatin A. Melosatin A (2 mg) was dissolved in MeOH (5 ml) and NABH₄ (2 mg) added slowly at room temp. After complete addition, the mixture was stirred at room temp for 3 hr. It was then diluted with water, extracted with CHCl₃, and the CHCl₃ layer washed twice with water. Removal of solvent left a colorless alcohol: m.p. 110–12°; MS *m/z* 355 (100, M⁺. C₂₁H₂₅NO₄), 321 (16, M⁺ - CH₃ - H₂O), 223 (24), 208 (48).

Methylation of 5-nitrovanillin. 5-Nitrovanillin (20 g) (Aldrich Chem. Co.) was dissolved in 50 ml MeOH and 100 ml ether, treated with 4 equivs diazomethane in 200 ml ether and left for 4 days.⁸ The solvent was then removed and the residue extracted with CHCl₃. The CHCl₃ layer was washed with 4% NaOH aq. to remove unreacted 5-nitrovanillin, then washed with water and dried over Na₂SO₄. Removal of solvent left a residue of 14 that crystallized from EtOH as needles: m.p. 89–90° (lit. m.p. 90–1°¹³); ¹H NMR 4.05, 4.12 (2s, 2 OCH₃, 6 H), 7.66, 7.86 (2d, *J* = 2 Hz, 2 ArH), 9.96 (s, CHO, 1 H); MS *m/z* 211 (100, M⁺. C₉H₉NO₅), 135 (31), 150 (20), 164 (26), 119 (23), 105 (29).

1-Phenyl-5-(3,4-dimethoxy-5-nitrophenyl)-1,4-pentadiene-3-one (16). Compound 14 (5.3 g) was dissolved in 150 ml EtOH and 3.65 g benzalacetone added. The mixture was cooled in an ice bath and 50 ml of 2% NaOH aq. added slowly with vigorous stirring. The stirring was continued for 3 hr whereupon 16 separated and was removed by filtration. (yield

4.69 g, 35%); m.p. 125–30°; MS m/z 339 (100, M^+ $C_{19}H_{17}NO_3$), 322 (37), 131 (40), 103 (45).

1-Phenyl-5-(3,4-dimethoxy-5-aminophenyl)pentan-3-ol hydrochloride (17). Compound 16 (2.39 g) was dissolved in 200 ml MeOH and 1 g NaBH₄ added slowly with stirring at room temp. The stirring was continued for 4 hr. The crude product was combined with 1 g of 10% Pd-C and hydrogenated for 5 hr. The catalyst was removed by filtration and the filtrate concentrated to dryness, diluted with 200 ml water, extracted twice with 200 ml CHCl₃ and the extract washed with water. The CHCl₃ extract was dried over Na₂SO₄ and the solvent evaporated. The residue was dissolved in 500 ml ether and treated with ethereal HCl. The hydrochloride of 17 was separated by filtration (0.684 g, 28% from 16); m.p. 180–5°; MS m/z 315 (39, M^+ $C_{19}H_{23}NO_3$), 180 (9), 167 (100), 154 (10), 152 (25), 91 (19).

2,3-Dimethoxy-5-(5-phenylpent-*x*-enyl)aniline hydrochloride. The derivative 17 recovered from its hydrochloride (600 mg) was dissolved in 500 ml dry benzene, 600 mg P₂O₅ added, and the mixture refluxed for 24 hr. The benzene was removed, and the residue diluted with water and neutralized with 5% Na₂CO₃. The free base was then extracted with CHCl₃ and the extracts washed with water, dried over Na₂SO₄ and the solvent evaporated. The residue was dissolved in 10 ml ether and precipitated with ethereal HCl. The crude hydrochloride (210 mg, 37%) melted at 170–5°; MS m/z 297 (54, M^+ $C_{19}H_{23}NO_2$), 206 (60), 167 (70), 166 (100), 152 (23), 91 (41).

Melosatin A (7). The above mixture of olefins (200 mg) was dissolved in 50 ml MeOH, 200 mg 10% Pd-C added, and hydrogenated for 4 hr. The catalyst was removed by filtration and the solvent evaporated leaving 100 mg (50%) of amorphous 18. This was treated with 1 ml oxalyl chloride and stirred at 165–70° for 2 hr. The reaction was monitored by tlc and a portion of the crude product purified by preparative tlc (R_f 0.61), CHCl₃–MeOH (95:5). The product was crystallized from benzene–petroleum ether and found identical in all respects (m.p., m.m.p., IR, UV, MS, ¹H NMR) with natural melosatin A.

4-Methylisatin. A soln of 9 g chloral hydrate in 120 ml water was combined with 130 g NaSO₄, 4.5 g *m*-toluidine in 30 ml water, 5.1 ml conc HCl and 11 g hydroxylamine hydrochloride in 50 ml water. This mixture was refluxed vigorously for two min and cooled, whereupon crystalline 3-methyl isonitrosoacetanilide separated (3.92 g, 53%); m.p. 142° (lit. m.p. 145°¹⁰). This product (3.5 g) was added slowly to 2 ml conc H₂SO₄ preheated to 60°. After complete addition, the temp was raised to 80° for 10 min. It was then cooled, poured onto ice, filtered, and the ppt washed with water. The crude 4-methylisatin was purified by dissolving it in 100 ml 8% NaOH aq. and carefully neutralizing the soln with dil HCl, and filtering precipitated impurities. The filtrate was then strongly acidified to pH 3 with dil HCl, precipitating the yellow 4-methylisatin which was filtered and recrystallized from EtOH (0.5 g, 16%); m.p. 185–8° (lit. m.p. 189°¹⁴); MS m/z 161 (63, M^+ $C_9H_7NO_2$), 133 (100), 106 (41), 105 (23), 104 (22), 103 (24); ¹H NMR δ 2.58 (s, ArCH₃, 3 H), 6.76 (d, J = 7 Hz, Ar5 H, 1 H), 6.88 (d, J = 7 Hz, Ar7 H, 1 H), 7.42 (t, J = 7 Hz, Ar6 H, 1 H), 8.60 (broad s, NH, 1 H).

4-Methyl-7-methoxyisatin. This compound was prepared by the method used above for 4-methylisatin, starting with 4.5 g 2-methoxy-5-methyl-aniline (Aldrich Chem. Co.). It gave a nearly quantitative yield of the isonitrosoacetanilide: m.p. 100–5°. Cyclization of 6 g of the isonitrosoacetanilide as above gave 3 g (54%) of red crystals of 4-methyl-7-methoxyisatin which were recrystallized from EtOH: m.p. 237–8° (lit. m.p. 235–6°¹⁵); MS m/z 191 (100, M^+ $C_{10}H_9NO_3$), 163 (48), 162 (39), 135 (77), 120 (30), 106 (30), 105 (28); ¹H NMR δ 2.52 (s, ArCH₃), 3.88 (s, OCH₃, 3 H), 6.79 (d, J = 8.5 Hz, Ar5 H, 1 H), 7.04 (d, J = 8.5 Hz, Ar6 H, 1 H), 7.75 (broad s, NH, 1 H).

4,7-Dimethoxyisatin. This compound was synthesized by the above method starting with 4.5 g of 2,5-dimethoxyaniline (Aldrich Chem. Co.). The isonitrosoacetanilide formed in

nearly quantitative yield: m.p. 120–5°; MS m/z 224 (100, M^+ $C_{10}H_{12}N_2O_4$), 209 (43), 193 (19), 179 (34), 164 (59), 138 (96). Cyclization of 1 g of this product and purification via tlc afforded 100 mg (11%) pure 4,7-dimethoxyisatin which was recrystallized from EtOH as red needles: m.p. 250–2°; MS m/z 207 (100, M^+ $C_{10}H_9NO_4$), 179 (79), 178 (43), 164 (28), 150 (43), 149 (31), 136 (31), 135 (31), 122 (50), 121 (32); ¹NMR δ 3.89 (s, OCH₃, 3 H), 3.91 (s, OCH₃, 3 H), 6.70 (d, J = 8.8 Hz, Ar5 H, 1 H), 7.36 (d, J = 8.8 Hz, Ar6 H, 1 H), 7.78 (broad s, NH, 1 H).

2,3-Dimethoxybenzamide. 2,3-Dimethoxybenzoic acid (5.5 g) was converted to the acid chloride by refluxing it in dry benzene with 3.6 g SOCl₂ and treating the crude product with conc NH₄OH. The crude amide was extracted into benzene, washed with bicarbonate, and the solvent evaporated. The residue recrystallized from benzene–petroleum ether giving the amide (2.39 g, 44%); m.p. 90° (lit. m.p. 93–4°C¹⁶).

2,3-Dimethoxyaniline hydrochloride. NaOH (2.4 g) in 20 ml water was combined with 0.6 ml Br₂ at 0°. Finely powdered 2,3-dimethoxybenzamide (1.81 g) was added and stirred until soln was complete whereupon it was heated at 70–80° for 30 min. Water (30 ml) was added and the aniline extracted with CHCl₃, the extracts washed with water, and the solvent evaporated. The residue was dissolved in ether and treated with HCl. The precipitated hydrochloride was then recrystallized from EtOH (1.15 g 61%); m.p. 193–4°. The picrate recrystallized from EtOH: m.p. 174–6°; lit. mp 173–5°¹⁷.

6,7-Dimethoxyisatin. 2,3-Dimethoxyaniline hydrochloride (0.5 g) was stirred with 1.26 g oxalyl chloride and the mixture then heated to 170° in an oil bath for 1.5 hr. The crude product was extracted with CHCl₃ and purified using preparative tlc with 5% MeOH in CHCl₃ (R_f 0.49). The yellow product crystallized from chloroform–petroleum ether as yellow needles (25 mg, 5%); m.p. 209–10° (lit. m.p. 212–3°¹⁸); MS m/z 207 (78, M^+ $C_{10}H_9NO_4$), 179 (30), 178 (33), 151 (76), 149 (100), 136 (20), 119 (20); ¹H NMR δ 3.91 (s, 7 OCH₃, 3 H), 3.98 (s, 6 OCH₃, 3 H), 6.59 (d, J = 8.36 Hz, Ar5 H, 1 H), 7.42 (d, J = 8.4 Hz, Ar4 H, 1 H), NH too broad to observe.

5-Methoxyisatin. Reddish-brown crystals (K&K Chemical Co.): ¹H NMR δ 3.84 (s, OCH₃, 3 H), 6.80 (d, J = 9.5 Hz, Ar 7 H, 1 H), 7.14 (m, Ar4 H and 6 H, 2 H), 7.60 (broad s, NH, 1 H).

5,6-Dimethoxyisatin. Reddish-brown crystals (K&K Chemical Co.): ¹H NMR δ 3.88 (s, Ar5 OCH₃, 3 H), 4.02 (s, Ar 6 OCH₃, 3 H), 6.52 (s, Ar 7 H, 1 H), 7.12 (s, Ar 4 H, 1 H), 8.20 (broad s, NH, 1 H).

Acknowledgements—Research at Howard University was supported by a National Institutes of Health Contract No. 1 CP 33266. We thank Dr. Julia F. Morton, University of Miami, Coral Gables, for arranging to provide the plant material used in this study and Mr. W. P. Maal, Curaçao, for collections.

REFERENCES

- G. J. Kapadia, B. D. Paul, J. V. Silvert, H. M. Fales and E. A. Sokoloski, *J. Am. Chem. Soc.* **97**, 6814 (1975).
- G. J. Kapadia, Y. N. Shukla, S. P. Basak, H. M. Fales and E. A. Sokoloski, *Phytochemistry* **17**, 1444 (1978).
- Y. N. Shukla, E. A. Sokoloski, H. M. Fales and G. J. Kapadia, *Ibid.*, **15**, 1788 (1976).
- G. J. Kapadia, Y. N. Shukla, J. F. Morton, and H. A. Lloyd, *Ibid.*, **16**, 1431 (1977).
- G. J. Kapadia, Y. N. Shukla, B. K. Chowdhury, S. P. Basak, H. M. Fales and E. A. Sokoloski, *J. Chem. Soc. Chem. Commun.* 535 (1977).
- R. Gompper, *Advances in Heterocyclic Chemistry* (Edited A. R. Katritzky, A. J. Boulton, and J. M. Logowski) Vol. 2, p. 245. Academic Press, New York (1963).
- Due to the small quantities available, these reactions were run on melosatin A only.
- We did not obtain any of the 5-nitro-3,4-dimethoxyacetophenone reported by O. L. Brady and L. B. Manjunath, *J. Chem. Soc.* **125**, 1060 (1924), even with 4 moles excess diazomethane.

- ⁹An attempt to catalytically hydrogenate **16** directly to **17** led to high molecular weight products in acid or neutral solvents, due, perhaps, to easy reduction of the nitro group to an amine followed by Michael addition to the unsaturated CO of another molecule.
- ¹⁰T. Sandmeyer, *Helv. Chim. Acta* **2**, 234 (1919).
- ¹¹E. Leete, *Biogenesis of Natural Compounds*, (Edited P. Bernfeld) p. 780. Macmillan, New York (1963).
- ¹²E. Medina and G. Spittler, *Chem. Ber.* **112**, 376 (1979).
- ¹³R. Pschorr and W. Stohrer, *Ber. Dtsch. Chem. Ges.* **35**, 4399 (1902).
- ¹⁴H. Wexler, *Chim.* **12**, 219 (1961); *Chem. Abstr.* **58**, 3379 (1963).
- ¹⁵H. Colombara, U.S. 1,856,210, 3 May (1932); *Chem. Abstr.* **26**, 3523 (1932).
- ¹⁶F. Mauthner, *J. Prak. Chem.* **112**, 64 (1926).
- ¹⁷G. S. Gibson, J. L. Simonsen, and M. G. Rau, *J. Chem. Soc.* **111**, 79 (1917).
- ¹⁸J. M. Gulland, R. Robinson, J. Scott and S. Thornley, *J. Chem. Soc.* 2924 (1929).